KITAZATO®

VITRIFICATION MEDIA

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EN-VITRIFICATION MEDIA

GENERAL INFORMATION Basic UDI-DI: 458223146VTUN This Instruction for Use covers: VT601, Tarset population: Patients undergoing ART procedures For professional use only. **INTENDED USE** This product is to be used for the vitrification of oocytes (MII) and embryos (hereafter referred as **specimen**). **CONTENT** • No. 0 (Color: vial) by Basic Solution (BS), 1×1.5 ml vial (only for oocyte (MII) vitrification) • No. 1 (Color: green) by Basic Solution Equilibrio (BS), 1×1.5 ml vial • No. 2 (Color: green) Vitrification solution (VS), 2×1.5 ml vials. **Caution:** Before use, check the specifications of the container and labelling (number on top of the cap, cap color, vial labelling color, solution name and volume). If there is any discrepancy in the information regarding the items mentioned above, don't use the product and contact the distributor. **Caution:** This product is sterilized. Handle it under aseptic conditions in a clean bench. **RECOMMENDED EQUIPMENT** Use a stereomicroscope for observation of specimens. Use a warming plate. Use devices intended for its particular use. We recommend the use of KITAZATO products in combination with this device. **COMPOSITION** • HEPES within Basic Medium Mixture (MEM) • Ethylene glycol • Dimethylsulfoxide (DMSO) • Trehalose • Hydroxypropylcellulose • Gentamicin (VT601K) **PRODUCT SPECIFICATIONS** • Sterility: SAL 10-3 • Endotoxin by LAL method(only • 1cel MEA (Blastocysts after 96h): ≥ 80% • pH (EP) • Osmolality (EP) Biocompatibility: Suitable for mouse embryo culture (EP) Storage: Store (EP) and can be downloaded from our website (https://kitazato-ivf.com) **INSTRUCTIONS FOR USE Preparation:** • Fill 90 % of the Cooling Rack with fresh liquid nitrogen. • Bring BS, VS and specimen to room temperature. **Caution:** Make sure that the cap with a suitable internal diameter for oocytes or embryos. The recommended internal diameters are 120 µm for oocytes (MII), 120 µm for proruclear stage embryos, 180 µm for cleavage stage embryos, and 280 µm for morula or blastocyst. **A. Vitrification procedure using Repro Plate** Equilibration: **Caution:** Equilibration procedures for Oocytes (MII) and Embryos are different. **Equilibration of oocytes (MII):** 1. Place 1 drop(s) (up to four together) in a 50 µL drop of BS on a sterile dish. 2. Dispense a 50 µL drop of BS close to the BS drop. Merge the two drops. Wait for 3 min. 3. Move oocyte(s) into a single 50 µL drop of BS and wash for 3 more drops. Wait for 3 min. 4. Move oocyte(s) into a single 50 µL drop of ES and wait for 6–9 min. **Equilibration of embryos:** 1. Place 2 embryo(s) (up to four together) in a 50 µL drop of BS on a sterile dish. 2. Dispense a 50 µL drop of BS close to the BS drop. Merge the embryos or 12–15 min for blastocysts. **Vitrification: Caution:** Complete steps from 1 to 6 between 60 and 90 seconds. 1. Aspirate the specimen from ES drop with the tip of a pipette. 2. Transfer the specimen to the VS2 drop for 30 seconds. 3. Aspirate the specimen with a pipette and transfer to a 50 µL drop of VS2 for 30 seconds. 4. Place the specimen near the black mark on the Cryotop® stage. 5. Load the specimen into the tip of the Cryotop® by capillary action. 6. Plunge the Cryotop® immediately in liquid nitrogen and shake it vigorously. 7. Insert the Cryotop® in the outer straw and store it in a liquid nitrogen tank. **Caution:** Complete steps 1 through 6 within 60 seconds (MII) or 15 minutes (Embryos). **Warming:** Warm the specimen in 1 ml of solution at 37°C for 1 minute. Aspirate the specimen and transfer to a 1 ml drop of TS for 3 min. Move the specimen to a 0.5 ml drop of DS for 3 min. Move the specimen to a 0.5 ml drop of WS for 5 min. Move the specimen to a 0.5 ml drop of WS for 1 min. Culture the specimen in a 0.5 ml drop of culture medium. **B. Vitrification procedure using standard dish** Equilibration: **Caution:** Equilibration procedures for Oocytes (MII) and Embryos are different. **Equilibration of oocytes (MII):** 1. Place oocyte(s) (up to four together) in a 50 µL drop of BS on a sterile dish. 2. Dispense a 50 µL drop of BS close to the BS drop. Merge the two drops. Wait for 3 min. 3. Move oocyte(s) into a single 50 µL drop of BS and wash for 3 more drops. Wait for 3 min. 4. Move oocyte(s) into a single 50 µL drop of ES and wait for 6–9 min. **Equilibration of embryos:** 1. Place embryo(s) (up to four together) in a 50 µL drop of BS on a sterile dish. 2. Dispense a 50 µL drop of BS close to the BS drop. Merge the embryos or 12–15 min for blastocysts. **Vitrification: Caution:** Complete steps from 1 to 6 between 60 and 90 seconds. 1. Aspirate the specimen from ES drop with the tip of a pipette. 2. Transfer the specimen to the VS2 drop for 30 seconds. 3. Aspirate the specimen with a pipette and transfer to a 50 µL drop of VS2 for 30 seconds. 4. Place the specimen near the black mark on the Cryotop® stage. 5. Load the specimen into the tip of the Cryotop® by capillary action. 6. Plunge the Cryotop® immediately in liquid nitrogen and shake it vigorously. 7. Insert the Cryotop® in the outer straw and store it in a liquid nitrogen tank. **Caution:** Complete steps 1 through 6 within 60 seconds (MII) or 15 minutes (Embryos). **Warming:** Warm the specimen in 1 ml of solution at 37°C for 1 minute. Aspirate the specimen and transfer to a 1 ml drop of TS for 3 min. Move the specimen to a 0.5 ml drop of DS for 3 min. Move the specimen to a 0.5 ml drop of WS for 5 min. Move the specimen to a 0.5 ml drop of WS for 1 min. Culture the specimen in a 0.5 ml drop of culture medium. **PRECAUTIONS** • Don't use the product if you notice any unusual regarding the specifications on the label (number, colour, name, volume). • Device is sterilized if the vial is unopened • Use sterilized equipment. • Don't use the product if you notice any unusual regarding the specifications on the label (number, colour, name, volume).

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